

Supporting Information for

Inactivation of the particulate methane monooxygenase (pMMO) in *Methylococcus capsulatus* (Bath) by acetylene[□]

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- III. **Oxidation of HCCH, CH₃CCH, and C₆H₅CCH mediated by a model trinuclear copper complex**

I. Supplementary Figures

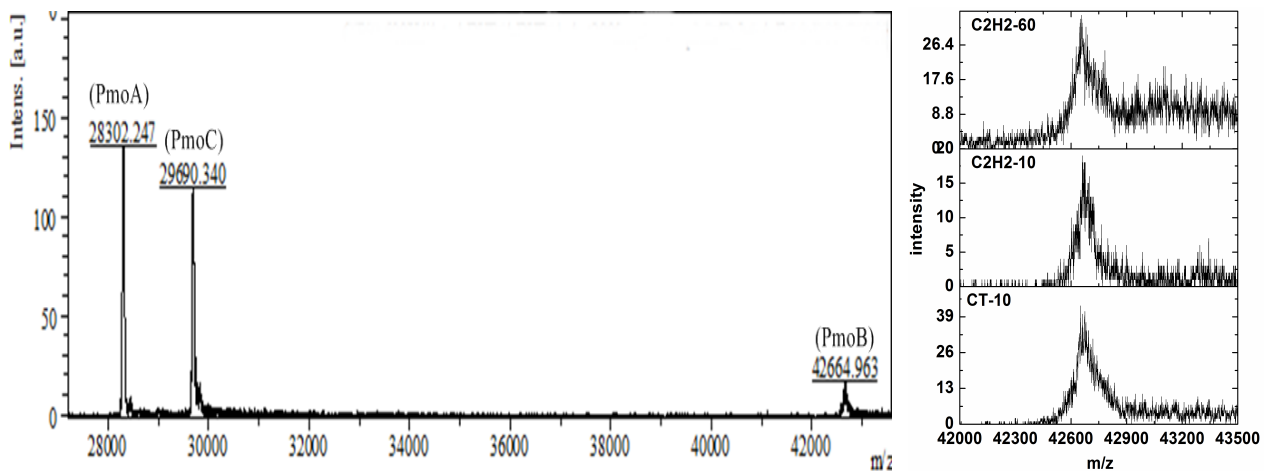


Fig. S1. *Left panel:* MALDI-TOF MS analysis of the intact pMMO complex without acetylene treatment; *Right panel:* Comparison of the mass peak of the PmoB subunit between untreated-sample (CT-10) and acetylene-treated samples at 10 min (C₂H₂-10) and 60 min (C₂H₂-60).

(A)	1	MHETKQGGEK	RFTGAICRCS	HRYSMEVKM	AAT	TIGGAAA	AEAPLLDKRW
	51	LTFALAIYTV	FYLWVRWYEG	VYGWSAGLDS	FAPEFETYWM	NFLYTEIVLE	
	101	IVTASILWGY	LWKTRDRNLA	ALTPREELRR	NFTHLVWLVA	YAWAIYWGAS	
	151	YFTEQDGTWH	QTIVRDTDFT	PSHIIEFYLS	YPIYIITGFA	AFIYAKTRLP	
	201	FFAKGISLPY	LVLVVGPFMI	LPNVGLNEWG	HTFWFMEELF	VAPLHYGFVI	
	251	FGWLALAVMG	TLTQTFYSFA	QGGLGQSLCE	AVDEGLIAK		
(B)	1	MSAAQSAVRS	HAEAVQVSRT	IDWMALFVVF	FVIVGSYHIH	AMLTMGDWDF	
	51	WSDWKDRRLW	VTVTPIVLVT	FPAAVQSYLW	ERYRLPWGAT	VCVLGLLLGE	
	101	WINRYFNFWG	WTYFPINFVF	PASLVPGAIL	LDTVLMLSGS	YLFTAIVGAM	
	151	GWGLIFYPGN	WPPIAPLHVP	VENNGMLMSI	ADIQGYNYVR	TGTPEYIRMV	
	201	EKGTLRTEGK	DVAPVSAFFS	AFMSILIIYFM	WHFIGRWFSN	ERFLQST	

Fig. S2. The LC-MS/MS analysis of the in-gel chymotryptic digestion of the “~27 kDa” and “~23 kDa” bands excised from the SDS-PAGE gel of the pMMO complex. The matched peptides are shown in red. (A) The sequence coverage of the “~27 kDa” protein band or PmoC is 65%, and (B) 74% for the “~23 kDa” protein band or PmoA.

PmoB	1 MKTIKDRIAK WSAIGLLSAV AATAFYAPSA SAHGEKSQAA FMRMRTIHWY 51 DLSWSKEKVK INETVEIKGK FHVFEQWPET VDEPDVAFLN VGMPPGVFIR 101 KESYIGGQLV PRSVRLKIGK TYDFRVVLKA RRPGDWHVHT MMNVQGGGPI 151 IGPGKWITVE GSMSEFRNPV TLTGQTVDL ENYNEGNTYF WHAFWFAIGV 201 AWIGYWSRRP IFIPRLMVD AGRADELVSA TDRKVAMGFL AATILIVVMA 251 MSSANSKYPI TIPLQAGTMR GMKPLELPAP TVSVKVEDAT YRVPGRAMRM 301 KLTITNHGNS PIRLGEFYTA SVRFLDSDVY KDTTGYPEDL LAEDGLSVSD 351 NSPLAPGETR TVDVTASDAA WEVYRLSDII YDPDSRFAGL LFFFDATGNR 401 QVVQIDAPLI PSFM
PmoC	1 MHETKQGGEK RFTGAICRCS HRYNSMEVKM AATTIGGAAA AEAPLLDKKW 51 LTFALAIYTV FYLWVRWYEG VYGWSAGLDS FAPEFETYWM NFLYTEIVLE 101 IVTASILWGY LWKTRDRNLA ALTPREELRR NFTHLWLVA YAWAIYWGAS 151 YFTEQDGTWH QTIVRDTDFT PSHIIEFYLS YPIYITGFAAFIYAKTRLP 201 FFAKGISLPY LVLVVGPFMI LPNVGLNEWG HTFWFMEELF VAPLHYGFVI 251 FGWLALAVMG TLTQTFYSFA QGGLGQSLCE AVDEGLIAK
PmoA	1 MSAAQSAVRS HAEAVQVSRT IDWMALFVVF FVIVGSYHIH AMLTMGDWDF 51 WSDWKDRRLW VTVTPMLVT FPAAVQSYLW ERYRLPWGAT VCVLGLLLGE 101 WINRYFNFVG WTYFPINFVF PASLVPGAIL LDTVLMLSGS YLFTAIVGAM 151 GWGLIFYPGN WPILAPLHVP VENNGMLMSI ADIQGYNYVR TGTPEYIRMV 201 EKGTLRTFGK DVAPVSAFFS AFMSILIYFM WHFIGRWFSN ERFLQST

Fig. S3. Sequence coverage in the LC-MS/MS analysis of the acetylene (HCCH)-treated pMMO complex (PmoA: green; PmoB: blue; and PmoC: purple). The amino acids that are not covered in the experiment are highlighted in red text.

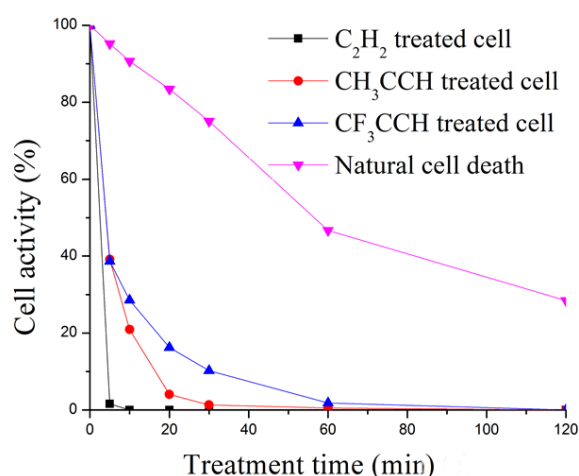


Fig. S4. Kinetics of inhibition of pMMO by three inhibitors based on whole cell assays. Cells of *M. capsulatus* (Bath) cultured under 30 μM Cu^{2+} are treated with HCCH, CH_3CCH , and CF_3CCH in separate experiments, and the activity of the pMMO is assayed by propylene oxidation at various times by GC-MS.

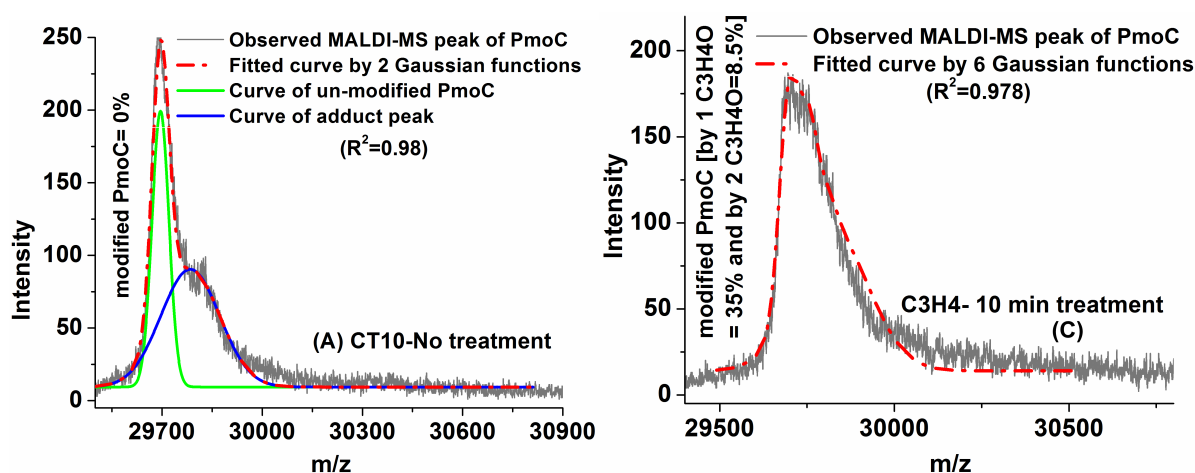


Fig. S5. Quantitation of the PmoC mass shift data. Fitting of the mass signals of the PmoC subunit from untreated and CH₃CCH-treated pMMO (C₃H₄-treatment) to a sum of contributions from chemically modified and unmodified species. The MALDI-TOF MS signal of PmoC after 10 min-treatment with C₃H₄ best fits 6 Gaussian functions representing signals from unmodified PmoC, PmoC modified with 1 methyl ketene, and PmoC modified with 2 methyl ketenes. For details of the fitting and calculation of the percentage of each chemical modification, see **Materials and Methods**.

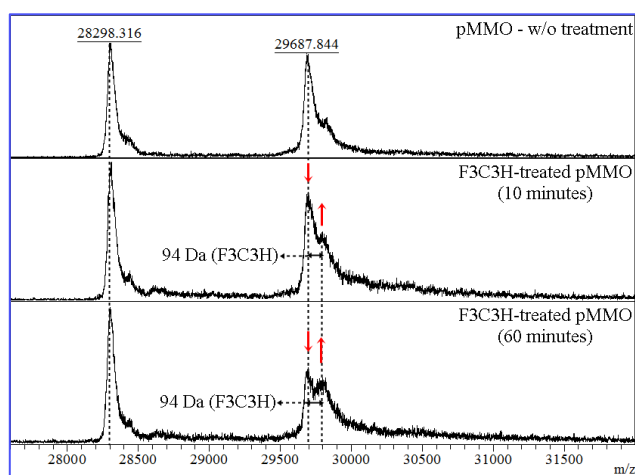


Fig. S6. MALDI-TOF MS analysis of the CF₃CCH-treated pMMO and untreated samples in time-course experiments. Whole cells were treated with and without CF₃CCH (F₃C₃H-treated pMMO) in separate experiments at two time points (10 and 60 min), as described in the kinetic study (Figure S4). Cells were then broken and the pMMO was purified. The MALDI-MS was performed directly on the intact purified pMMO proteins (see **Materials and Methods** section for details).

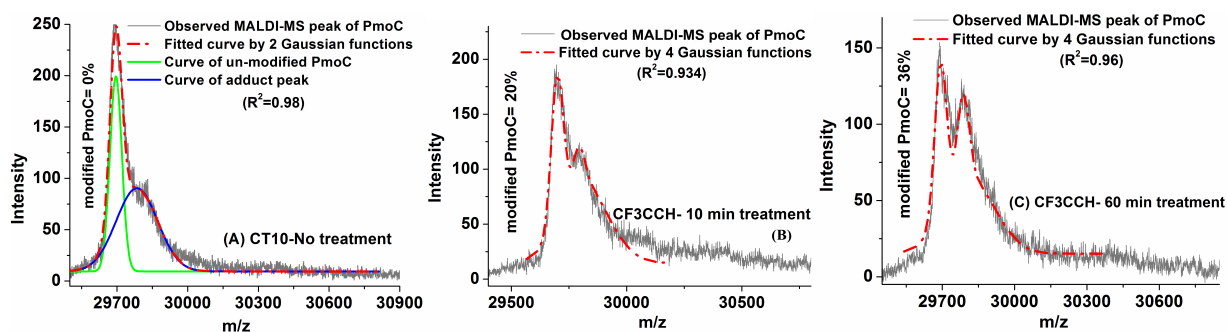


Fig. S7. Quantitation of the PmoC mass shift data. Fitting of the mass signals of the PmoC subunits from the untreated and CF₃CCH-treated pMMO to a sum of contributions from chemically modified and unmodified species. The percentage of chemically modified PmoC subunit was determined from the relative contribution of the two species to the composite PmoC mass signal in each case: *panel A*, untreated pMMO; *panel B*, CF₃CCH-treated pMMO (10 min); and (*panel C*), CF₃CCH-treated pMMO (60 min). Two Gaussian functions and four Gaussian functions were used to fit the unmodified PmoC subunit and the alkyne-modified PmoC subunit, respectively. For details of the fitting, see **Materials and Methods**.

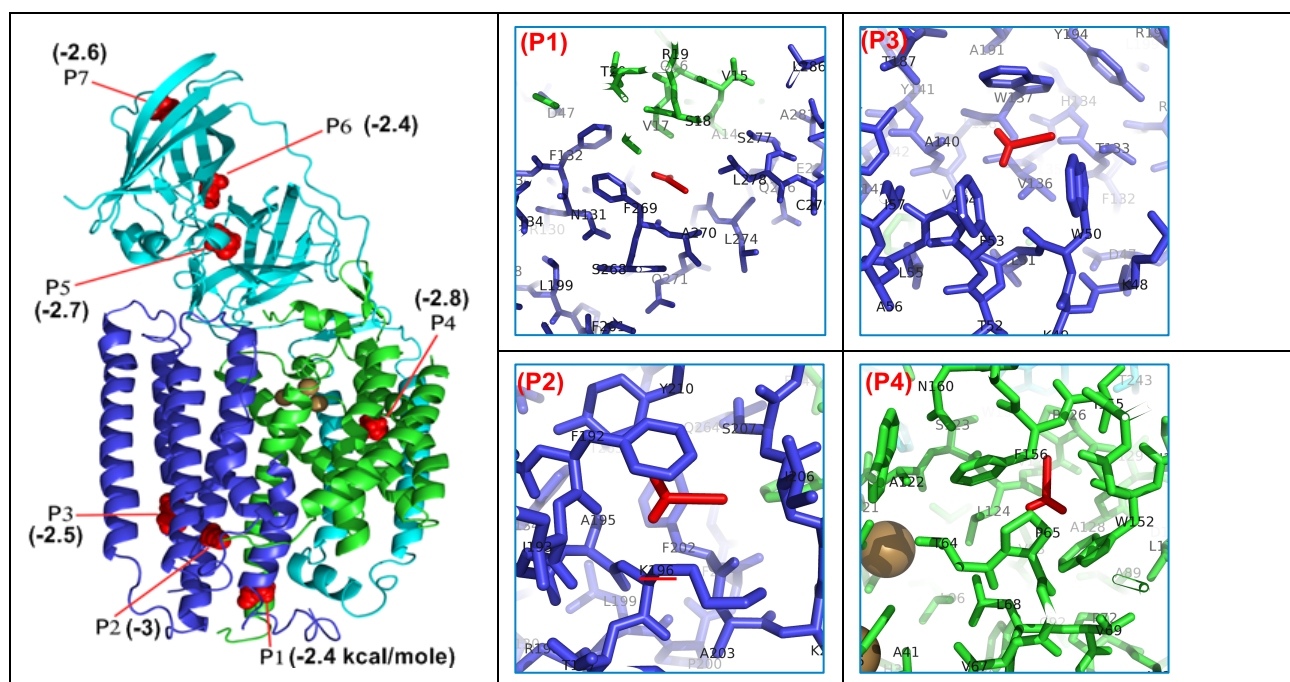
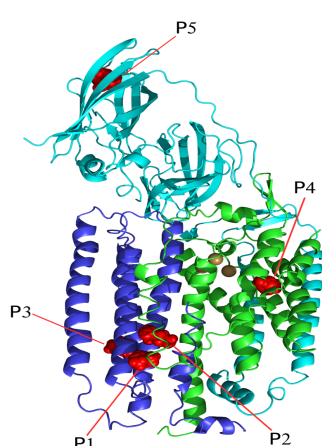


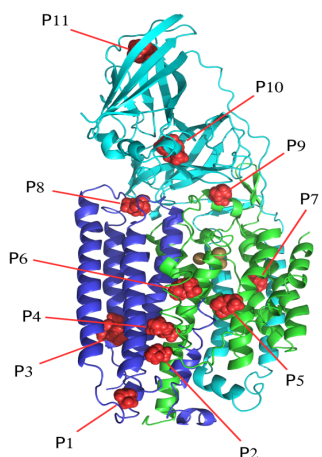
Fig. S8. Molecular docking of ketene to the revised crystal structure of pMMO (accession number 3RGB). There are 7 affinity binding sites of ketene denoted from P1 to P7. The surrounding residues of four affinity sites within the transmembrane domain are presented in the right. Predicted binding energy (kcal/mole) of ketene to each position is provided in parentheses, showing the highest binding affinity for P2. K196 is underlined in (P2) showing its proximity to the affinity binding site number 2 or P2, which is also in the cluster of two other sites P1 and P3. The putative tricopper cluster (D site) is highlighted in

brown spheres. Crystal structure of pMMO from *M. capsulatus* used in this study is reproduced from PDB (accession number 3RGB).



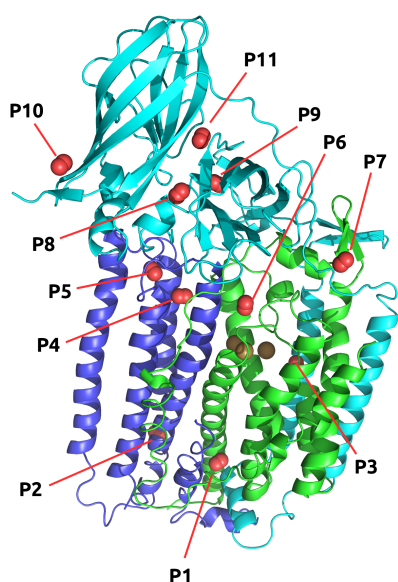
	CH ₄	C ₂ H ₆	C ₃ H ₈	C ₄ H ₁₀	C ₅ H ₁₂	C ₂ H ₂
P1	-1.5	-2.9	-3.8	-3.8	×	-2.9
P2	-1.3	-2.5	-3.3	-3.8	-4.3	-2.4
P3	-1.2	-2.2	-2.9	-3.5	-4.0	-2.3
P4	-1.2	-2.3	-3.1	-3.7	-4.2	-2.3
P5	-1.1	-2.1	-3.0	-3.7	-4.0	-2.2

Fig. S9. Predicted binding sites of *n*-alkanes (C1-C5) as well as the suicide-substrate acetylene by molecular dockings in the crystal structure of pMMO (accession number 1YEW) and the corresponding binding energies (kcal/mole). × indicates no binding affinity.



	Methanol	Ethanol	Propan-2-ol	Butan-2-ol	Pentan-2-ol
P1	-1.9	-2.6	-3.0	×	×
P2	×	-2.8	×3.0	×	×
P3	×	×	×	-3.6	-3.8
P4	-2.0	-3.0	×	×	-3.9
P5	×	×	-3.2	-3.6	-4.0
P6	×	×	-3.2	-3.5	-3.8
P7	-1.9	-2.7	-3.4	-4.1	-4.3
P8	-2.0	-2.7	-3.0	-3.4	×
P9	-1.9	×	×	-3.4	-3.8
P10	-1.9	-2.6	-3.1	-3.5	-3.9
P11	-2.1	-3.0	-3.7	-4.5	-4.5

Fig. S10. Predicted binding sites of some alcohols in pMMO and the corresponding binding energies (kcal/mole). × indicates no binding affinity.



Positions	Binding energy (units)
P1	-2.1
P2	-2.0
P3	-2.2
P4	-2.1
P5	-2.0
P6	-2.1
P7	-2.0
P8	-2.1
P9	-2.4
P10	-2.0
P11	-2.1

Fig. S11. Predicted binding sites of dioxygen in pMMO and the corresponding binding energies (kcal/mole).

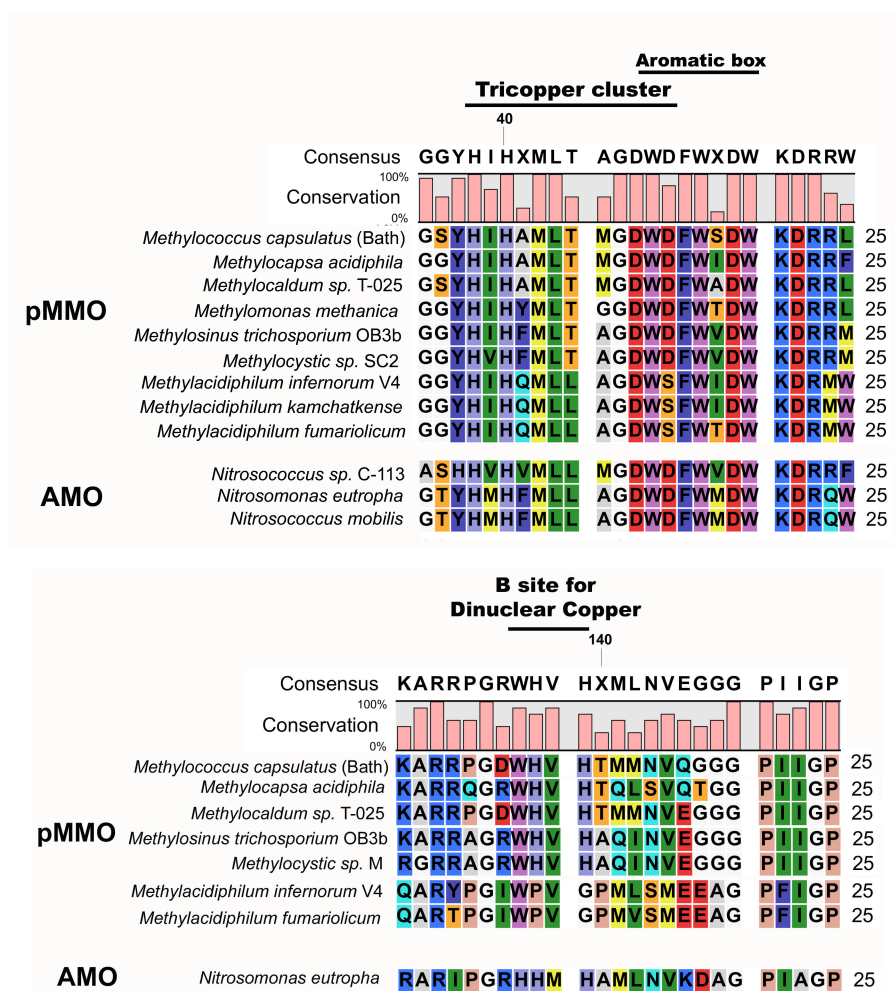


Fig. S12. Upper panel: Sequence alignments of the PmoA subunits in the region containing both the copper-binding residues for the tricopper cluster and the substrate-binding pocket (aromatic box) in pMMOs from proteobacterial methanotrophs and extremely acidophilic methanotrophs. A similar putative copper-binding peptide domain has been identified in several ammonia monooxygenases. **Lower panel:** Sequence alignments of the PmoB subunits in the region containing the copper-binding residues for the dinuclear B site in pMMOs from proteobacterial methanotrophs and the corresponding region in the PmoB subunits of extremely acidophilic methanotrophs. These binding residues are not conserved between the two types of methanotrophs. However, a similar copper-binding peptide domain has been identified in the AMO from *Nitrosomonas eutropha*.

Fig. S13. Whole sequence alignments between AMO and pMMO for subunit A (PmoA vs AmoA) and subunit C (PmoC vs AmoC) using ClustalW.

Whole sequence alignment of PmoA vs AmoA

```

>_ Pmo-A                                     247 aa vs.
>_ Amo-A                                     276 aa

scoring matrix: , gap penalties: -12/-2
44.6% identity;          Global alignment score: 849

-
      10      20      30      40      50
Pmo-A  MSAAQS-----AVRSHAEAVQVSRTIDWMALFVVFFVIVGSYHIHAMLTMGDWDFWSDWK
      ::  ..      ::      ::      ::      ::      ::      ::      ::
Amo-A  MSIFRTEEILKAAKMPPEAVHMSRLIDAVYFPILIILLVGTYHMHFMLLAGDWDFFWMDWK
      10      20      30      40      50      60

      60      70      80      90      100     110
Pmo-A  DRRLWVTVTPIVLVTFPAAVQSYLWERYRLPWGATVCVLGLLLGEWINRYFNFVGWTFYFP
      ::  :  :::::  ::  ....  ::  ::  :::::  :::::  :::::  :::::
Amo-A  DRQWWPVVTPIVGITYCSAIMYYLWVNYRQPFGATLCVVCLLIGEWLTRYWGFYWWSHYP
      70      80      90      100     110     120

      120     130     140     150     160     170
Pmo-A  INFVFPASLVPGAIIIDTVLMLSGSYLFTAIVGAMGWGLIFYPGNWPPIIAPLHVPVEYNG
      ::::  :  :::::  ::::  ..  :::::  :::::  :::::  :::::  ::::  ::
Amo-A  INFVTPGIMLPALMLDFTLYLTRNLVTALVGGGFFGLLFYPGNWPPIFGPTHLPVVEG
      130     140     150     160     170     180

      180     190     200     210     220     230
Pmo-A  MLMSIADIQGYNYVRTGTPEYIRMVEKGTLRFTFGKDVPVSAFFSAFMSILIIYFMWHFIG
      :::::  ::  :::::  :::::  :::::  ..  :::::  :::::  ::  ::
Amo-A  TLLSMADYMGHLYVRTGTPEYVRHIEQGSRLRTFGGHTTVIAAFFSAFVSMLMFTVWWYLG
      190     200     210     220     230     240

      240
Pmo-A  RW-----FSNERFLQST-
      .      :::  :  ..
Amo-A  KVYCTAFFYVKGKRGRIVHRNDVTAFGEEGFPEGIK
      250     260     270

```

Whole sequence alignment of PmoC vs AmoC

>_ Pmo-C 289 aa vs.

>_ Amo-C 271 aa

scoring matrix: , gap penalties: -12/-2

40.5% identity; Global alignment score: 768

```

      10      20      30      40      50
PmoC  MHETKQGGEKRFTGAICRCSHRYNSMEVVKMAATTIGGAAAAEAPLLDKKWLTFAL-AIYT
      :  :  :  :..  :  :  :.  :  :  :...  :..  :.
AmoC  M-ATTLG-----TSSASSVSSRGYDMSLWY-----DSKFYKFGMITMLL
              10      20      30

      60      70      80      90     100     110
PmoC  VFYLWVRWYEGVYGWSAGLDSFAPEFETYWMNPLYTEIVLEIVTASILWGYLWKTRD--R
      :  .:  :.  :..  :.:  :.:  :.:  :.:  :..  :.  :.  :.:  :.:  :.
AmoC  VAIFWV-WYQRYFAYSHGMDSMEPEFDRVWMGLWRVHMAIMPLFALVTWGWILKTRDTKE
      40      50      60      70      80      90

      120     130     140     150     160     170
PmoC  NLAALTPREELRRNFTHLVWLVAWAIYWGASYFTEQDGTWHQTIVRDTDFTPSHIIEF
      :  :  :.  :.:  :.  :.:  :.  :.:  :.:  :.:  :.:  :.:  :.:  :.
AmoC  QLDNLDPKLEIKRYFYMMWLGVYIFGVYWGGSFTEQDASWHQVIIRDTSFTPSHVVMF
      100     110     120     130     140     150

      180     190     200     210     220     230
PmoC  YLSYPIYIITGFAAFIYAKTRLPPFFAKGISLPLVLVVGPFMILPNVGLNEWGHTFWFME
      :  :.:  :.  :.:  :.:  :.:  :.:  :.:  :.:  :.:  :.:  :.:  :.:
AmoC  YGSFPMYIVCGVATYLYAMTRLPLFSRGISFPLVMAIAGPLMILPNVGLNEWGHAFWFME
      160     170     180     190     200     210

      240     250     260     270     280
PmoC  ELFVAPLHYGFVIFGWLALAVMGTLTQ--TFYSFAQGGLGQSLCEAVDEGLIAK
      :.:  :.:  :.:  :.  :.  :.  :.:  :.  :.  :.  :.  :.
AmoC  ELFSAPLHWGFVVLGWAGLFQGGVAAQIIITRYSNLTDVVWNNQSKEILNNRIVA
      220     230     240     250     260     270

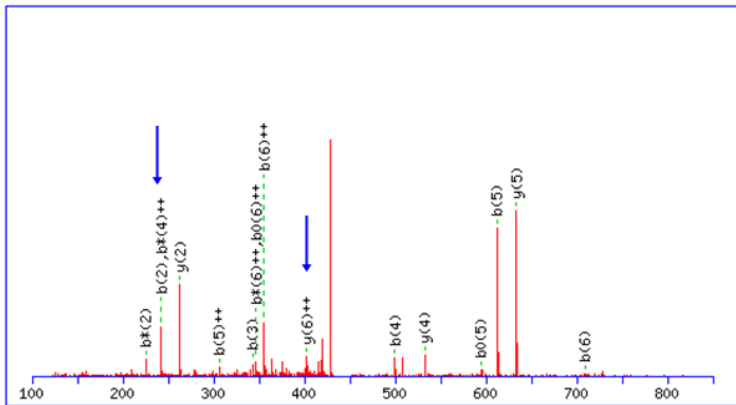
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II. Supplementary Tables

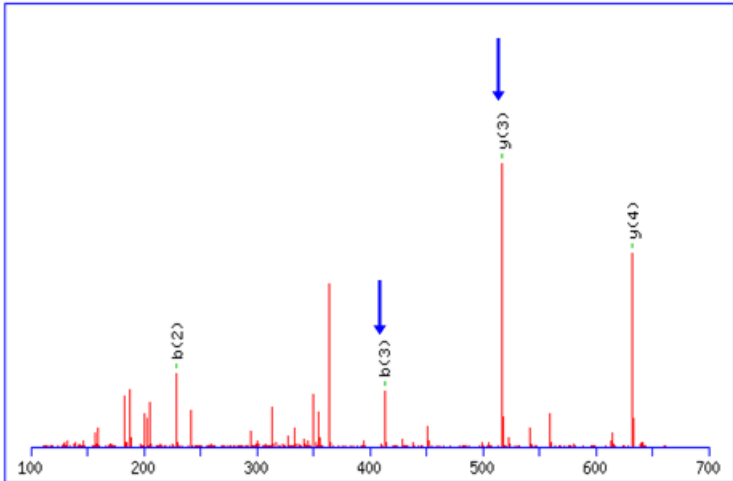
Table S1. Predicted molecular masses of the pMMO subunits and the values in Daltons measured by MALDI-TOF MS.

Subunit	Predicted Molecular mass (Average)	Observed Molecular mass (Average)	NCBI reference sequence
PmoB	42 649.000	42 664.663	YP_114234.1
PmoC	29 689.360	29 690.340	YP_114236.1
PmoA	28 294.240	28 302.247	YP_114235.1

Table S2. MS/MS spectra of the identified peptides labeled by ketene (C_2H_2O , +42.0106 Da), methyl ketene (C_3H_4O , +56.0262 Da) and trifluoropropyne (F_3C_3H , +94.003 Da). The blue arrows illustrate the b and/or y ions from the fragments of the identified peptides.

Residue #	Chemical modification	MS/MS spectra of identified peptides																																																																																
K196 (PmoC)	Ketene (C ₂ H ₂ O, +42.0106 Da)	<div><div><div>Query Observed Mr(expt) Mr(calc) Delta Miss Score Expect Rank Peptide Modification</div><div><div><div>✓</div><div>690</div><div>437.7614</div><div>873.5082</div><div>873.5072</div><div>1.18</div><div>0</div><div>(21)</div><div>0.0078</div><div>1</div><div>Y.AKTRLP.F + C2H2O-</div><div></div></div></div></div><div></div><div><p>Monoisotopic mass of neutral peptide Mr(calc): 873.5072</p><p>Variable modifications:</p><p>K2 : C2H2O-KSR (KRS)</p><p>Ions Score: 21 Expect: 0.0012</p><p>Matches (Bold Red): 16/52 fragment ions using 30 most intense peaks</p></div><table><thead><tr><th>#</th><th>b</th><th>b⁺⁺</th><th>b[*]</th><th>b⁺⁺⁺</th><th>b⁰</th><th>b⁰⁺⁺</th><th>Seq.</th><th>y</th><th>y⁺⁺</th></tr></thead><tbody><tr><td>1</td><td>72.0444</td><td>36.5258</td><td></td><td></td><td></td><td></td><td>A</td><td></td><td></td></tr><tr><td>2</td><td>242.1499</td><td>121.5786</td><td>225.1234</td><td>113.0653</td><td></td><td></td><td>K</td><td>803.4774</td><td>402.24</td></tr><tr><td>3</td><td>343.1976</td><td>172.1024</td><td>326.1710</td><td>163.5892</td><td>325.1870</td><td>163.0972</td><td>T</td><td>633.3719</td><td>317.18</td></tr><tr><td>4</td><td>499.2987</td><td>250.1530</td><td>482.2722</td><td>241.6397</td><td>481.2881</td><td>241.1477</td><td>R</td><td>532.3242</td><td>266.66</td></tr><tr><td>5</td><td>612.3828</td><td>306.6950</td><td>595.3562</td><td>298.1817</td><td>594.3722</td><td>297.6897</td><td>L</td><td>376.2231</td><td>188.61</td></tr><tr><td>6</td><td>709.4355</td><td>355.2214</td><td>692.4090</td><td>346.7081</td><td>691.4250</td><td>346.2161</td><td>P</td><td>263.1390</td><td>132.07</td></tr><tr><td>7</td><td></td><td></td><td></td><td></td><td></td><td></td><td>F</td><td>166.0863</td><td>83.54</td></tr></tbody></table></div>	#	b	b ⁺⁺	b [*]	b ⁺⁺⁺	b ⁰	b ⁰⁺⁺	Seq.	y	y ⁺⁺	1	72.0444	36.5258					A			2	242.1499	121.5786	225.1234	113.0653			K	803.4774	402.24	3	343.1976	172.1024	326.1710	163.5892	325.1870	163.0972	T	633.3719	317.18	4	499.2987	250.1530	482.2722	241.6397	481.2881	241.1477	R	532.3242	266.66	5	612.3828	306.6950	595.3562	298.1817	594.3722	297.6897	L	376.2231	188.61	6	709.4355	355.2214	692.4090	346.7081	691.4250	346.2161	P	263.1390	132.07	7							F	166.0863	83.54
#	b	b ⁺⁺	b [*]	b ⁺⁺⁺	b ⁰	b ⁰⁺⁺	Seq.	y	y ⁺⁺																																																																									
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6	709.4355	355.2214	692.4090	346.7081	691.4250	346.2161	P	263.1390	132.07																																																																									
7							F	166.0863	83.54																																																																									
K196 (PmoC)	Methyl ketene (C ₃ H ₄ O, +56.0262 Da)	<div><div><div>✓</div><div>794</div><div>444.7686</div><div>887.5226</div><div>887.5229</div><div>-0.25</div><div>0</div><div>32</div><div>0.0007</div><div>1</div><div>Y.AKTRLP.F + C3H4O-</div><div></div></div></div>																																																																																

		<div></div> <p>Monoisotopic mass of neutral peptide Mr(calc): 887.5229 Variable modifications: K2 : C3H4O-KSR (KRS) Ions Score: 32 Expect: 0.00014 Matches (Bold Red): 12/52 fragment ions using 23 most intense peaks</p> <table><thead><tr><th>#</th><th>b</th><th>b⁺⁺</th><th>b[*]</th><th>b^{*++}</th><th>b⁰</th><th>b⁰⁺⁺</th><th>Seq.</th><th>y</th><th>y⁺⁺</th></tr></thead><tbody><tr><td>1</td><td>72.0444</td><td>36.5258</td><td></td><td></td><td></td><td></td><td>A</td><td></td><td></td></tr><tr><td>2</td><td>256.1656</td><td>128.5864</td><td>239.1390</td><td>120.0731</td><td></td><td></td><td>K</td><td>817.4931</td><td>409.2502</td></tr><tr><td>3</td><td>357.2132</td><td>179.1103</td><td>340.1867</td><td>170.5970</td><td>339.2027</td><td>170.1050</td><td>T</td><td>633.3719</td><td>317.1896</td></tr><tr><td>4</td><td>513.3144</td><td>257.1608</td><td>496.2878</td><td>248.6475</td><td>495.3038</td><td>248.1555</td><td>R</td><td>532.3242</td><td>266.6657</td></tr><tr><td>5</td><td>626.3984</td><td>313.7028</td><td>609.3719</td><td>305.1896</td><td>608.3879</td><td>304.6976</td><td>L</td><td>376.2231</td><td>188.6152</td></tr><tr><td>6</td><td>723.4512</td><td>362.2292</td><td>706.4246</td><td>353.7160</td><td>705.4406</td><td>353.2239</td><td>P</td><td>263.1390</td><td>132.0731</td></tr><tr><td>7</td><td></td><td></td><td></td><td></td><td></td><td></td><td>F</td><td>166.0863</td><td>83.5468</td></tr></tbody></table>	#	b	b ⁺⁺	b [*]	b ^{*++}	b ⁰	b ⁰⁺⁺	Seq.	y	y ⁺⁺	1	72.0444	36.5258					A			2	256.1656	128.5864	239.1390	120.0731			K	817.4931	409.2502	3	357.2132	179.1103	340.1867	170.5970	339.2027	170.1050	T	633.3719	317.1896	4	513.3144	257.1608	496.2878	248.6475	495.3038	248.1555	R	532.3242	266.6657	5	626.3984	313.7028	609.3719	305.1896	608.3879	304.6976	L	376.2231	188.6152	6	723.4512	362.2292	706.4246	353.7160	705.4406	353.2239	P	263.1390	132.0731	7							F	166.0863	83.5468
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K48 (PmoC)	Methyl ketene (C ₃ H ₄ O, +56.0262 Da)	<input checked="" type="checkbox"/> <u>416</u> 373.2156 744.4166 744.4170 -0.48 1 (19) 0.014 1 L.LD K KW.L + C3H4O																																																																																

		<div></div> <p>Monoisotopic mass of neutral peptide Mr(calc): 744.4170 Variable modifications: K3 : C3H4O-KSR (KRS) Ions Score: 19 Expect: 0.011 Matches (Bold Red): 4/34 fragment ions using 6 most intense peaks</p> <table><thead><tr><th>#</th><th>b</th><th>b⁺⁺</th><th>b⁺</th><th>b⁺⁺⁺</th><th>b⁰</th><th>b⁰⁺⁺</th><th>Seq.</th><th>y</th><th>y⁺</th></tr></thead><tbody><tr><td>1</td><td>114.0913</td><td>57.5493</td><td></td><td></td><td></td><td></td><td>L</td><td></td><td></td></tr><tr><td>2</td><td>229.1183</td><td>115.0628</td><td></td><td></td><td>211.1077</td><td>106.0575</td><td>D</td><td>632.3402</td><td>316.6</td></tr><tr><td>3</td><td>413.2395</td><td>207.1234</td><td>396.2129</td><td>198.6101</td><td>395.2289</td><td>198.1181</td><td>K</td><td>517.3133</td><td>259.1</td></tr><tr><td>4</td><td>541.3344</td><td>271.1709</td><td>524.3079</td><td>262.6576</td><td>523.3239</td><td>262.1656</td><td>K</td><td>333.1921</td><td>167.0</td></tr><tr><td>5</td><td></td><td></td><td></td><td></td><td></td><td></td><td>W</td><td>205.0972</td><td>103.0</td></tr></tbody></table>	#	b	b ⁺⁺	b ⁺	b ⁺⁺⁺	b ⁰	b ⁰⁺⁺	Seq.	y	y ⁺	1	114.0913	57.5493					L			2	229.1183	115.0628			211.1077	106.0575	D	632.3402	316.6	3	413.2395	207.1234	396.2129	198.6101	395.2289	198.1181	K	517.3133	259.1	4	541.3344	271.1709	524.3079	262.6576	523.3239	262.1656	K	333.1921	167.0	5							W	205.0972	103.0
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K49 (PmoC)	Methyl ketene (C ₃ H ₄ O, +56.0262 Da)	<div><input checked="" type="checkbox"/> 415 373.2155 744.4164 744.4170 -0.75 1 22 0.0065 1 L.LDKKWL + C3H4O</div>																																																												

III. Oxidation of HCCH, CH₃CCH, and C₆H₅CCH mediated by a model trinuclear copper complex.

If the oxidation of hydrocarbons by pMMO is mediated by the putative tricopper cluster at site D of the enzyme, as has been implicated by a broad range of biochemical/biophysical studies on the enzyme [1, 2] as well as oxidation of methane and other hydrocarbons recently demonstrated for biomimics of the tricopper cluster [3, 4], then the same tricopper complex should be able to oxidize the suicide substrates HCCH and CH₃CCH to their corresponding ketene derivatives. To confirm this chemistry, we have compared the oxidation of the two suicide substrates with the hydrocarbon substrates of pMMO mediated by the [Cu^ICu^ICu^I(**7-Ethppz**)]¹⁺ tricopper complex, where **7-Ethppz** stands for the ligand 3,3'-(1,4-diazepane-1,4-diyl)bis(1-(4-ethylhomopiperazin-1-yl)propan-2-ol) [5]. Recently, Chan and coworkers have demonstrated that the related [Cu^ICu^ICu^I(**7-N-Etppz**)]¹⁺ complex, where **7-N-Etppz** stands for the ligand 3,3'-(1,4-diazepane-1,4-diyl)bis[1-(4-ethylpiperazine-1-yl)propan-2-ol], is capable of catalyzing the efficient oxidation of methane and many other straight-chain hydrocarbons from C₂-C₆ at room temperature, when the complex is activated by O₂ or H₂O₂ [3, 4].

For HCCH and CH₃CCH, we have confirmed that they are indeed converted into ketene products by the model tricopper complex. Products of the oxidation reactions are analyzed by GC-MS as shown in Figure SIII.1 and Figure SIII.2. Although there is no ketene intermediate observed directly by GC-MS, the compounds identified are products of the reactions between the expected ketene intermediate with oxygen and water. This is not surprising as the ketene intermediates are very reactive species, which will quickly react with O₂ or H₂O within the reaction buffer. For instance: (i) ketene reacts with O₂ to form CO₂ and formaldehyde (HCHO); and (ii) the methyl ketene reacts with O₂ to form CO₂ and acetaldehyde (CH₃CHO), or with H₂O to give acetic acid (CH₃COOH) [6-9]. All these products are identified by GC-MS.

To bolster our understanding of the mechanism of the oxidation of HCCH and CH₃CCH mediated by the model tricopper complex, we have also employed ethynyl-benzene (C₆H₅CCH, or C₈H₆) as a substrate to take advantage of its well-known ketene chemistry [8]. The addition of an “O-atom” across the triple bond of this alkyne will promote cyclization to yield the benzofuran and acetophenone. Indeed, the benzofuran is observed. In addition, there are also other products coming from the reactions of the oxidation product of ethynyl-benzene intermediate with O₂ and H₂O. As a control, we have observed no reaction between O₂ and this alkyne without the tricopper catalyst, indicating that the

formation of the ketene derivative is indeed mediated by the tricopper cluster at room temperature.

A proposed mechanism for the oxidation of alkynes catalyzed by the $[\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}(\text{7-Ethppz})]^{1+}$ complex upon activation by H_2O_2 is given in Figure SIII.3.

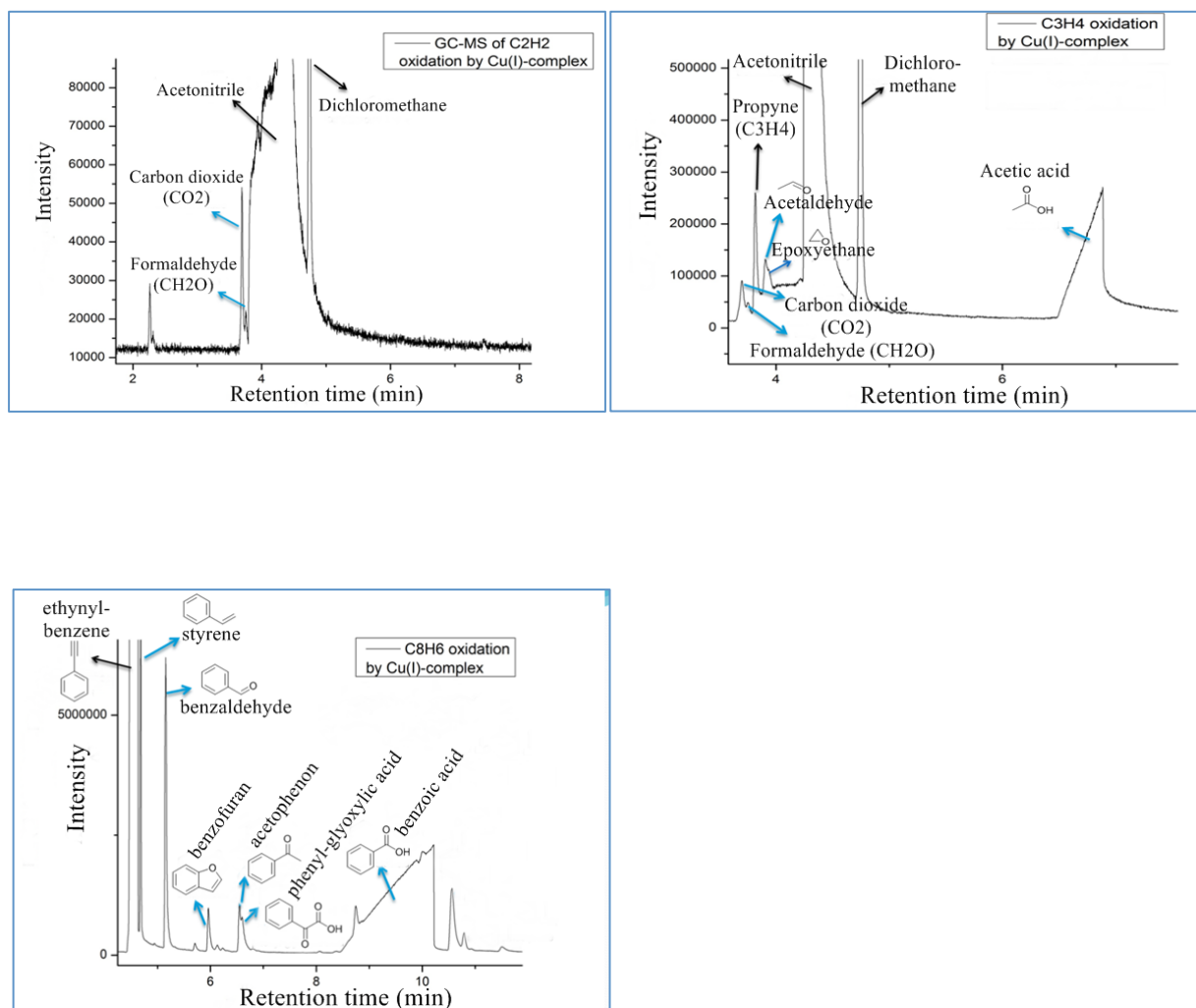
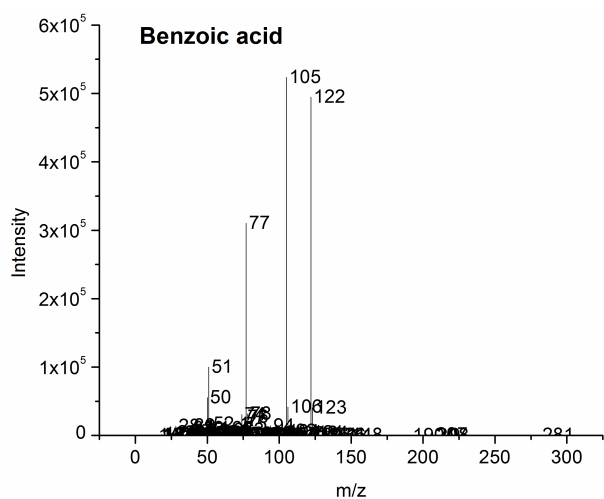
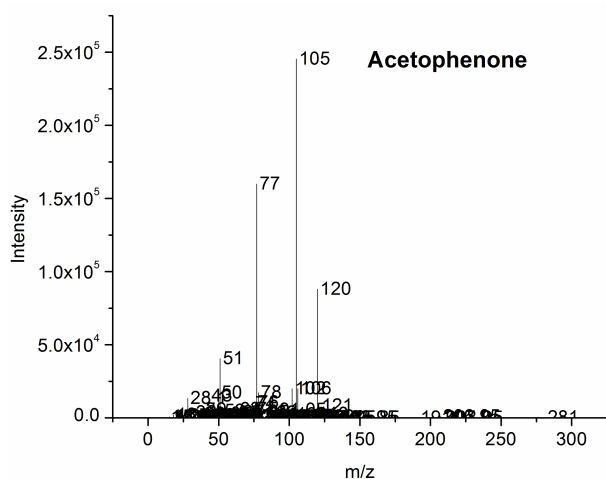
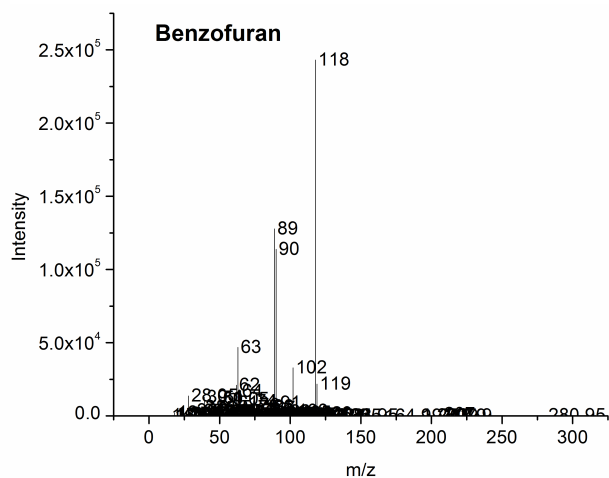
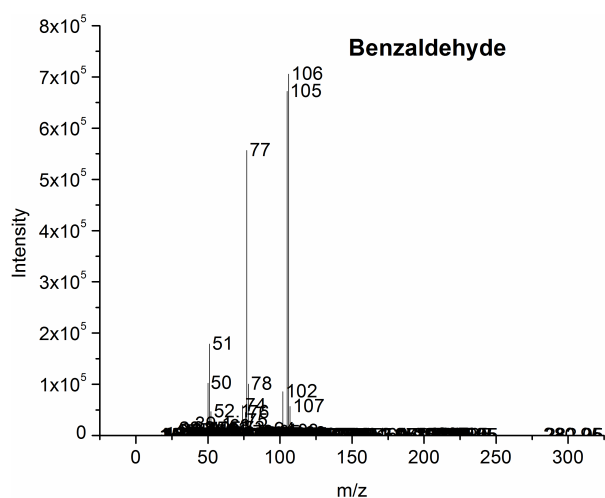
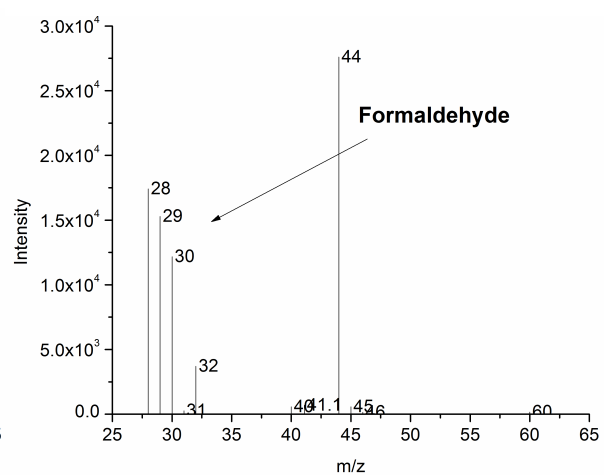
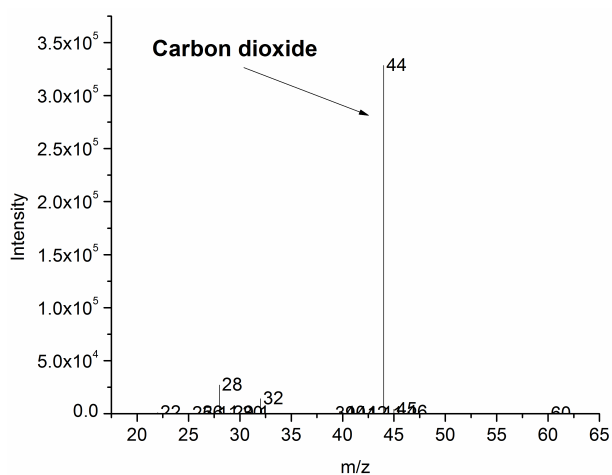


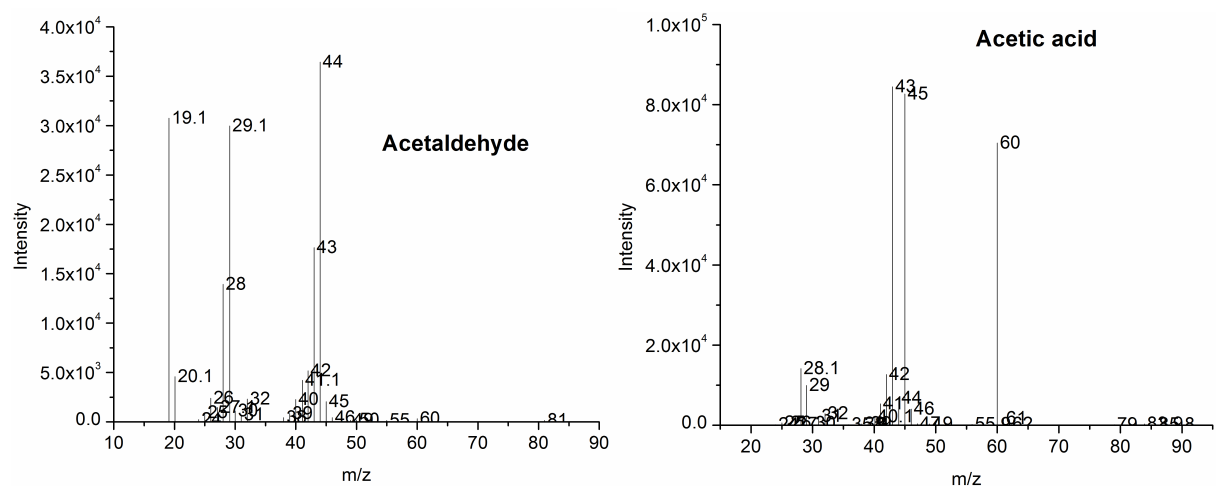
Fig SIII.1. GC-MS analysis of the oxidation of alkynes catalyzed by $[\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}(\text{7-Ethppz})]^{1+}$ complexes mediated by H_2O_2 . The black and blue arrows highlight peaks of the solvent and the products that are formed by the reactions, respectively.

+ MS spectrum of the products of ethynyl-benzene (C_8H_6) oxidation



+ MS spectrum of the products of CH_3CCH oxidation





+ MS spectrum of the products of HCCH oxidation

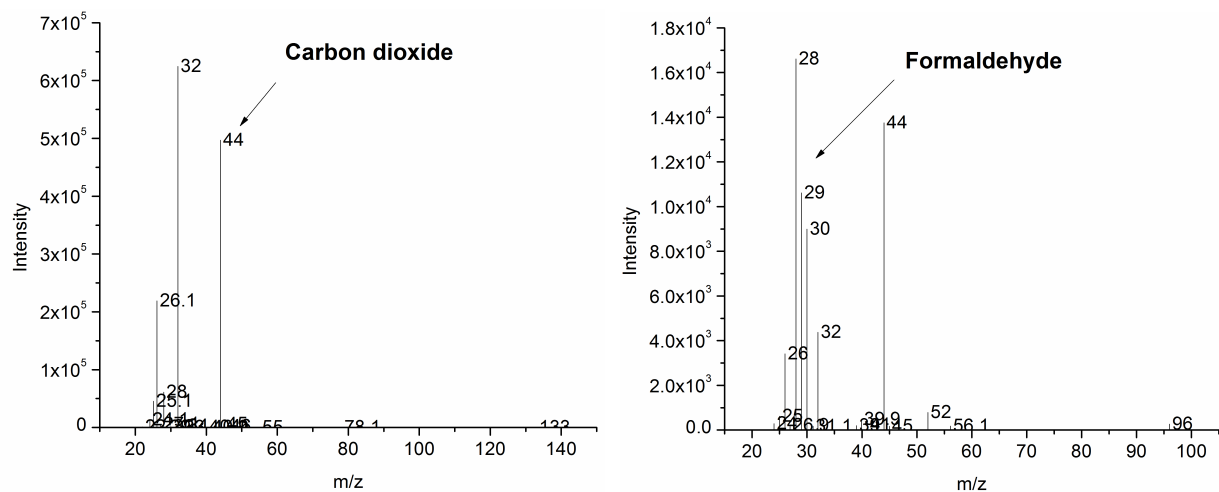


Fig SIII.2. Mass spectrum of products identified from the oxidation of alkynes catalyzed by $[\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}(\text{7-Ethppz})]^{\text{I}+}$ complexes mediated by H_2O_2 .

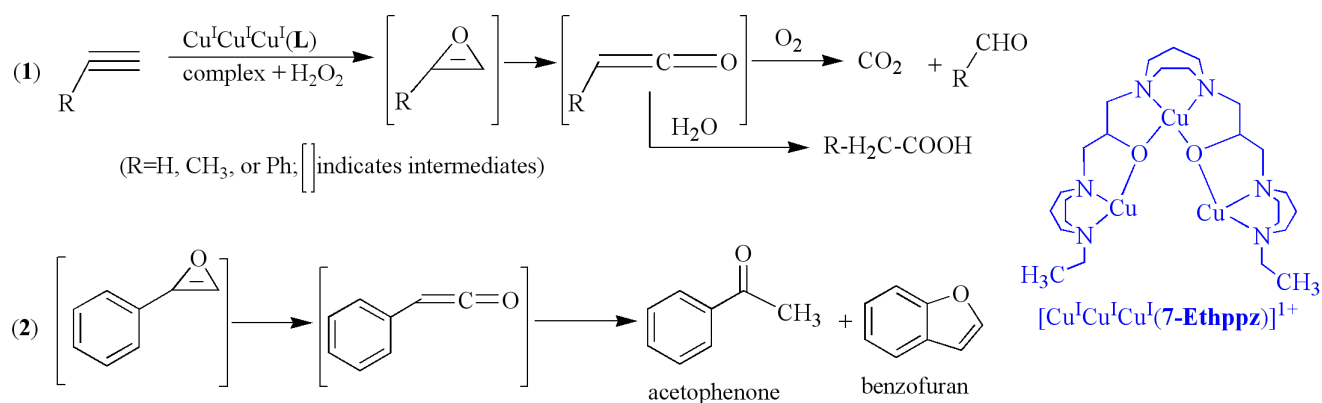


Fig SIII.3. A proposed mechanism for the oxidation of alkynes catalyzed by the $[\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}(\text{7-Ethppz})]^{1+}$ complex upon activation by H_2O_2 . All reactions are conducted at room temperature, and the products are analyzed by GC-MS. Reaction (1) applies to all three alkynes: HCCH , CH_3CCH , and ethynyl-benzene ($\text{C}_6\text{H}_5\text{CCH}$). Reaction (2) is applicable to ethynyl-benzene only. The inorganic complex to the right represents structure of the $[\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}\text{Cu}^{\text{I}}(\text{7-Ethppz})]^{1+}$ tricopper cluster selected for the study.

References

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